

Clinical Research Article

Clinical and Hormonal Profiles Correlate With Molecular Characteristics in Patients With 11 β -Hydroxylase Deficiency

Melek Yildiz,^{1,2} Emregul Isik,³ Zehra Yavas Abali,⁴ Mehmet Keskin,⁵ Mehmet Nuri Ozbek,⁶ Firdevs Bas,² Seyit Ahmet Ucakturk,⁷ Muammer Buyukinan,⁸ Hasan Onal,¹ Cengiz Kara,⁹ Karl-Heinz Storbeck,¹⁰ Feyza Darendeliler,² Atilla Cayir,¹¹ Edip Unal,⁶ Ahmet Anik,¹² Huseyin Demirbilek,¹³ Tugba Cetin,¹⁴ Fatma Dursun,¹⁵ Gonul Catli,¹⁶ Serap Turan,⁴ Henrik Falhammar,¹⁷ Tugba Baris,¹⁸ Ali Yaman,¹⁹ Goncagul Haklar,¹⁹ Abdullah Bereket,⁴ and Tulay Guran⁴

¹Department of Pediatric Endocrinology, Kanuni Sultan Suleyman Training and Research Hospital, Istanbul, Turkey; ²Department of Pediatric Endocrinology and Diabetes, Istanbul University, Istanbul Faculty of Medicine, Istanbul, Turkey; ³Clinics of Pediatric Endocrinology, Gaziantep Children's Hospital, Gaziantep, Turkey; ⁴Department of Pediatric Endocrinology and Diabetes, Marmara University, Faculty of Medicine, Istanbul, Turkey; ⁵Department of Pediatric Endocrinology and Diabetes, Gaziantep University, School of Medicine, Gaziantep, Turkey; ⁶Department of Pediatric Endocrinology and Diabetes, SBU Diyarbakir Gazi Yasargil Education and Research Hospital, Diyarbakir, Turkey; ⁷Department of Pediatric Endocrinology, Ankara City Hospital, Children's Hospital, Ankara, Turkey; ⁸Department of Pediatric Endocrinology, Konya Training and Research Hospital, Konya, Turkey; ⁹Department of Pediatrics, Division of Pediatric Endocrinology, Altinbas University, Faculty of Medicine, Istanbul, Turkey; ¹⁰Department of Biochemistry, Stellenbosch University, Stellenbosch, South Africa; ¹¹Department of Pediatric Endocrinology and Diabetes, Erzurum Training and Research Hospital, Erzurum, Turkey; ¹²Department of Pediatric Endocrinology and Diabetes, Adnan Menderes University, School of Medicine, Aydin, Turkey; ¹³Department of Pediatric Endocrinology and Diabetes, Hacettepe University, School of Medicine, Ankara, Turkey; ¹⁴Department of Pediatric Endocrinology, Sanliurfa Training and Research Hospital, Sanliurfa, Turkey; ¹⁵Department of Pediatric Endocrinology and Diabetes, Istanbul University of Health Science, Umraniye Training and Research Hospital, Istanbul, Turkey; ¹⁶Department of Pediatric Endocrinology, Izmir Katip Celebi University, School of Medicine, Izmir, Turkey; ¹⁷Department of Endocrinology, Metabolism and Diabetes, Karolinska University Hospital, Stockholm, Sweden; Department of Molecular Medicine and Surgery, Karolinska Institutet, Stockholm, Sweden; ¹⁸Gelisim Genetik Tani Merkezi, Istanbul, Turkey; and ¹⁹Department of Biochemistry, Marmara University, Faculty of Medicine, Istanbul, Turkey

ORCID numbers: 0000-0002-6603-2983 (M. Yildiz); 0000-0003-1669-6383 (K.-H. Storbeck); 0000-0001-6374-5884 (H. Demirbilek); 0000-0002-5622-6987 (H. Falhammar); 0000-0003-2658-6866 (T. Guran).

Received: 26 November 2020; Editorial Decision: 1 April 2021; First Published Online: 8 April 2021; Corrected and Typeset: 16 July 2021.

Abstract

Background: Given the rarity of 11 β -hydroxylase deficiency (11 β OHD), there is a paucity of data about the differences in clinical and biochemical characteristics of classic (C-11 β OHD) and nonclassic 11 β OHD (NC-11 β OHD).

Objective: To characterize a multicenter pediatric cohort with 11 β OHD.

Method: The clinical and biochemical characteristics were retrospectively retrieved. *CYP11B1* gene sequencing was performed. Seventeen plasma steroids were quantified by liquid chromatography-mass spectrometry and compared to that of controls.

Results: 102 patients (C-11 β OHD, n = 92; NC-11 β OHD, n = 10) from 76 families (46,XX; n = 53) had biallelic *CYP11B1* mutations (novel 9 out of 30). Five 46,XX patients (10%) were raised as males. Nineteen patients (19%) had initially been misdiagnosed with 21-hydroxylase deficiency. Female adult height was 152 cm [−1.85 SD score (SDS)] and male 160.4 cm (−2.56 SDS). None of the NC-11 β OHD girls had ambiguous genitalia (C-11 β OHD 100%), and none of the NC-11 β OHD patients were hypertensive (C-11 β OHD 50%). Compared to NC-11 β OHD, C-11 β OHD patients were diagnosed earlier (1.33 vs 6.9 years; $P < 0.0001$), had higher bone age-to-chronological age ($P = 0.04$) and lower adult height (−2.46 vs −1.32 SDS; $P = 0.05$). The concentrations of 11-oxygenated androgens and 21-deoxycortisol were low in all patients. The baseline ACTH and stimulated cortisol were normal in NC-11 β OHD. Baseline cortisol; cortisone; 11-deoxycortisol; 11-deoxycorticosterone and corticosterone concentrations; and 11-deoxycortisol/cortisol, 11-deoxycorticosterone/cortisol, and androstenedione/cortisol ratios were higher in C-11 β OHD than NC-11 β OHD patients ($P < 0.05$). The 11-deoxycortisol/cortisol ratio >2.2 , <1.5 , and <0.1 had 100% specificity to segregate C-11 β OHD, NC-11 β OHD, and control groups.

Conclusion: NC-11 β OHD can escape from clinical attention due to relatively mild clinical presentation. However, steroid profiles enable the diagnosis, differential diagnosis, and subtyping of 11 β OHD.

Key Words: *CYP11B1*, congenital adrenal hyperplasia, steroid profiling, 11-oxygenated androgens, adrenal insufficiency, androgen excess, children

Steroid 11 β -hydroxylase deficiency (11 β OHD) (OMIM#202010) caused by biallelic mutations of *CYP11B1* is the second most common type of congenital adrenal hyperplasia (CAH) and accounts for 0.2% to 8% of all CAH cases (1-3). 11 β OHD occurs in ~1:100 000 to 1:200 000 live births in general nonconsanguineous populations (3,4). However, we recently reported a higher incidence of 1:60 000 in the Turkish population (5).

The metabolic signature of 11 β OHD includes raised concentrations of adrenocorticotropic hormone (ACTH), 11-deoxycortisol, 11-deoxycorticosterone (DOC), and adrenal androgen precursors together with low cortisol, corticosterone, and aldosterone. The clinical manifestations of cortisol deficiency are not apparent due to the glucocorticoid effect of high DOC concentrations, which also activates the mineralocorticoid receptor, leading to hypertension (2,3). In addition, an increased steroidogenic flux toward adrenal androgen precursors results in hyperandrogenemia. 11 β OHD can present as classic (C-11 β OHD) or nonclassic

(NC-11 β OHD) phenotypes depending on the degree of clinical severity and percentage loss of *CYP11B1* activity (3). The clinical features of C-11 β OHD include hyporeninemic hypokalemic hypertension, virilization of female external genitalia (46,XX difference/disorder of sex development), precocious pseudopuberty, accelerated skeletal maturation and short adult height. The frequency of NC-11 β OHD is even rarer than the C-11 β OHD, and only few genetically confirmed NC-11 β OHD cases have been reported (6-13). NC-11 β OHD is characterized by normal external genitalia at birth and can manifest later in life with milder virilization, precocious pseudopuberty, hirsutism, or menstrual irregularities, while arterial hypertension is less frequently observed (3).

Given the rarity of 11 β OHD, there is a paucity of data about the differences in clinical and biochemical characteristics of C- and NC-11 β OHD. In this study, we report a national cohort of 102 children with 11 β OHD with extensive phenotyping, steroid profiling by mass spectrometry,

molecular genetic analysis of the *CYP11B1* gene and clinical data to study the differences between the patients with C- and NC-11 β OHD. The aim was to expand our knowledge on the clinical, biochemical and molecular spectrum of 11 β OHD.

Methods

Study Population

We performed a cross-sectional study recruiting children with clinical and biochemical features suggestive of 11 β OHD from 16 Departments of Pediatric Endocrinology across Turkey. The medical records of the patients were reviewed for detailed clinical and biochemical information, in addition to other relevant medical and family history data.

Excess plasma samples from children examined for other conditions, such as well-controlled type 1 diabetes, euthyroid hypothyroidism or hyperthyroidism on treatment or simple growth retardation but without adrenal enzyme deficiency, polycystic ovary syndrome, or puberty disorder, were used as controls for adrenal steroid measurements. The control group included 210 children [119 girls and 91 boys, median age 11.4 (0.8-19.5) years]. Age and gender of control subjects were not different from the 11 β OHD patients ($P = 0.98$ and $P = 0.78$, respectively).

The study was performed with the approval of the Ethics Committee of Marmara University Faculty of Medicine, Istanbul, Turkey (09.2017.476). Patients and/or parents provided written informed consent.

Hormonal Assays

Blood samples were collected between 8:00 and 9:00 AM after 48 h off-treatment state in a hospital setting with continuous observation for general well-being, blood pressure, and the other vital signs. These procedures were performed after obtaining an informed consent from the patient and/or the parents. We did not observe any adverse events in the patients during the off-treatment period. Biochemical assessment included adrenal steroid profile measured by liquid chromatography-mass spectrometry (LC-MS/MS) as previously described (14). ACTH and other biochemical parameters were measured using commercial immunoassays.

Molecular Genetic Analysis of the *CYP11B1* Gene

Genomic DNA was extracted using a QIAamp DNA Mini Kit (QIAGEN, Germantown, MD, USA) from peripheral blood lymphocytes according to standard protocols. The coding exons and exon-intron boundaries of the *CYP11B1*

gene (NM_000497) were amplified as previously described (15), sequenced via Sanger sequencing using an ABI 3130XL DNA Sequencer (Thermo Fisher Scientific, Waltham, MA, USA), and analyzed by Seqscape sequencing analysis software, version 2.7 (Applied Biosystems, Foster City, CA, USA). Sequence variants were designated according to Human Genome Variation Society recommendations (www.hgvs.org/rec.html) using the reference sequences GenBank NC_000008 (g.DNA), NM_000497 (c.DNA), and NP_000488.3 (protein).

Patients were classified as C- or NC-11 β OHD depending on the degree of clinical severity and percentage loss of *CYP11B1* activity as reported previously (6-13). The differences between C- and NC-11 β OHD were reviewed, focusing specifically on the clinical parameters and adrenal steroid profiles.

Statistical analysis

Statistical analysis was performed using GraphPad Prism® V5.0 software (GraphPad Software Inc, San Diego, California, USA). Data were tested for normality distribution using the Shapiro-Wilk test. Results are reported as frequencies and percentages, median with minimum-maximum, interquartile ranges (IQR) or 95% confidence intervals (95%CI) as appropriate. A nonparametric t test was used for comparison of numeric and χ^2 test for categorical variables. Statistical significance was set at $P < 0.05$.

Results

Clinical Characteristics of the Patients

We identified 102 patients (53 with 46,XX and 49 with 46,XY karyotypes) from 76 families (45 familial cases from 19 families and 57 singletons) diagnosed with 11 β OHD and genetically confirmed using *CYP11B1* gene mutation analysis. The patients had been assessed from the neonatal period until 20.8 years of age (median: 9.9 years). The age at diagnosis and initiation of hydrocortisone treatment was 1.8 years (range: 0.01 to 15.8). Eighty patients (78%) were diagnosed before 4 years of age, 35 of 53 girls (66%) were presented before 1 year of age because of ambiguous genitalia, while 43 of 49 boys (87%) presented after 1 year of age, mainly due to precocious pseudopuberty and advanced growth (Fig. 1A). Thus, the girls were diagnosed earlier than the boys [0.04 (range: 0.01 to 4.5) vs 2 (range: 0.04 to 12) years, $P < 0.0001$]. Twenty-nine patients (28%) had hyperpigmentation, and 10 patients (10%) had mild hyponatremia at the presentation (sodium levels 131-134 mEq/L). Six of them erroneously used fludrocortisone. Five 46,XX

patients (10%) were raised as males. Nineteen patients (19%) had been initially misdiagnosed as 21-hydroxylase deficiency (21OHD). Premature adrenarche was present in 51 (50%) patients, and 46 patients (46%) required treatment with at least 1 antihypertensive drug for hypertension (1 [range: 1 to 3] antihypertensive drugs). The age of the onset of antihypertensive therapy was 7.7 (range:1.5 to 16) years. The patients had markedly advanced bone age in both sexes [bone age/chronological age; females: 2.1 (range:0.80 to 15.00), $n = 15$ vs males: 2.4 (range:1.0 to 8.2), $n = 37$; $P = 0.67$] (Fig. 1B). Nineteen patients used gonadotropin-releasing hormone analogue in addition to hydrocortisone treatment to improve adult height. Adult height, adult height SD score (SDS), and corrected height SDS (adult height SDS – midparental SDS) were 152 (range: 136 to 162.5) cm, -1.85 (range: -4.62 to -0.10), and -1.07 (range: -3.45 to 0.79), respectively, for females ($n = 18$) and 160.4 (range: 134.2 to 175.3) cm, -2.56 (range: -6.81 to -0.15), and -1.85 (range: -4.87 to 1.64), respectively, for males ($n = 24$; $P = 0.22$ and $P = 0.11$ for adult height SDS and corrected height SDS, respectively). Fifteen of 49 boys (30%) had bilateral adrenal rest tumors [age at first detection 10 years (1.5 to 16)]. Detailed clinical features of the patients are provided in Supplemental Table 1 in (16).

Molecular Analysis of *CYP11B1* Gene

Thirty sequence variations were identified including 15 missense (novel $n = 5$: p.Glu310Gln, p.His69Pro, p.Gly446Asp, p.Phe487Cys, p.Arg412Leu), 3 nonsense, 7 splice-site (novel $n = 2$:c.1122-1G>C, c.955-3C>A) changes; 1 intronic change (novel $n = 1$: c.239 + 150C>A); 3 small deletions (novel $n = 1$:p.Glu198del); and 1 small insertion and 1 small duplication (Table 1). Ninety-four patients were homozygous and 8 were compound heterozygous for *CYP11B1* mutations. The parents carried the mutant alleles in the heterozygous state.

All of the novel single nucleotide variants were predicted to be “damaging,” “pathogenic,” or “disease-causing” using multiple in silico analysis tools including Polyphen-2, SIFT, Mutationtaster and PROVEAN (29-32). None of the variants were found in the population databases GnomAD (Genome Aggregation Database), ExAC, 6500ESP, or 1000 Genomes (33).

The c.1179_1180dup (p.Asn394Argfs*37), c.896T>C (p.Leu299Pro), c.954G>A (p.Thr318=) and c.421C>T (p.Arg141*) were the most common mutations accounting for two thirds of the alleles with allele frequencies of 24.7%, 16%, 10%, and 7.2%, respectively. Ten patients were subsequently classified as NC-11 β OHD based on the previous

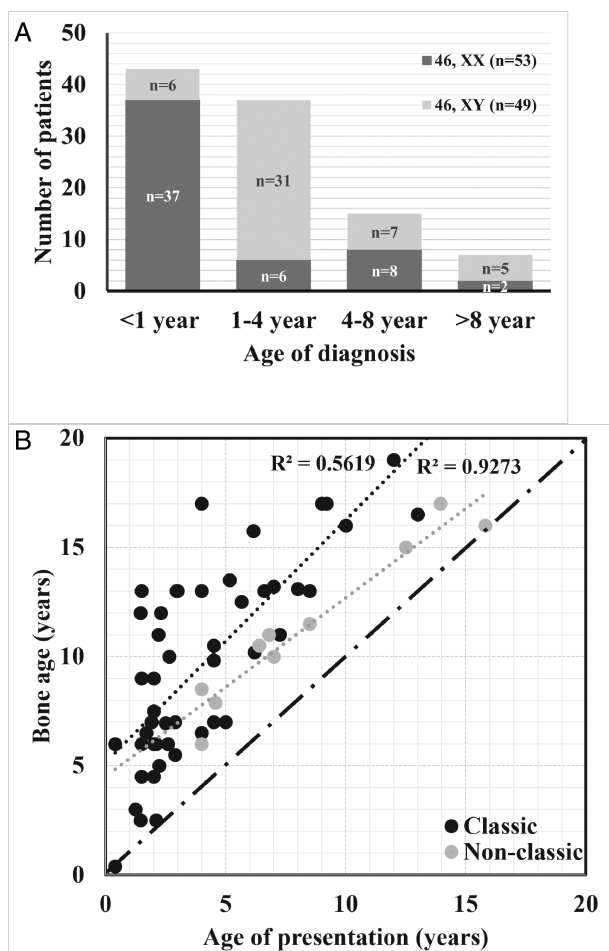


Figure 1. Clinical profile of 102 pediatric patients with 11 β -hydroxylase deficiency. (A) The age at diagnosis in female and male patients with 11 β -hydroxylase deficiency. Eighty patients (78%) were diagnosed before 4 years of age, 37 of 53 girls (70%) presented before 1 year of age because of ambiguous genitalia, while 43 of 49 boys (87%) presented after 1 year, mainly due to precocious pseudopuberty and advanced growth. (B) Bone age vs chronological age in classic (black) and nonclassic (grey) 11 β -hydroxylase deficiency patients at the presentation. The black dotted line represents a slope of 1 (bone age = chronological age). Almost all patients showed advanced bone age, being more pronounced in patients with classic 11 β -hydroxylase deficiency.

reports of mutations associated with NC-11 β OHD in the literature (6-13).

Comparisons Between C-11 β OHD and NC-11 β OHD

The median ages of the 92 C-11 β OHD (46,XX $n = 45$, 46,XY $n = 47$) and 10 NC-11 β OHD (46,XX $n = 8$, 46,XY $n = 2$) patients at the time of assessment were 9.8 (range: 0.07 to 20.8) and 10.4 (range: 4.66 to 18.8) years, respectively ($P = 0.93$). None of the NC-11 β OHD girls had ambiguous genitalia (C-11 β OHD 100%, $P < 0.0001$), and none of the NC-11 β OHD patients were hypertensive (C-11 β OHD 50%, $P = 0.0019$). The C-11 β OHD patients

Table 1. Characteristics, distribution, and the effect of CYP11B1 mutations in 102 patients with 11 β -hydroxylase deficiency

| # | Nucleotide change (genome assembly GRCh37:p13, CYP11B1 (NM_000497)) | Protein | Alleles (n) | Homozygous patients (n) | Heterozygous patients (n) | Localization | Activity (%) | Clinical phenotype | Reference | MAF in GnomAD (%) | RefSNPs (rs) |
|----|---|------------------|-------------|-------------------------|---------------------------|---------------|--------------|--------------------|------------|-------------------|--------------|
| 1 | c.1179_1180dup | p.Asn394Argfs*37 | 51 | 24 | 3 | e7/K-L loop | ND | C | 17 | 0.0008 | rs758714890 |
| 2 | c.896T>C | p.Leu299Pro | 33 | 15 | 3 | e5/H-I loop | 1 | C | 18 | 0.0004 | rs387907573 |
| 3 | c.954G>A | p.Thr318= | 21 | 10 | 1 | e5/I helix | ND | C | 19 | 0.0012 | rs753774484 |
| 4 | c.421C>T | p.Arg141* | 15 | 7 | 1 | e3/C helix | ND | C | 20 | 0.0008 | rs775479837 |
| 5 | c.928G>C | p.Glu310Gln | 14 | 7 | 0 | e5/I helix | ND | C | This study | NA | |
| 6 | c.372delG | p.His125Thrfs*8 | 12 | 6 | 0 | e2/B-C loop | ND | C | 21 | NA | rs1554653520 |
| 7 | c.1342C>T | p.Arg448Cys | 4 | 2 | 0 | e8/Cys pocket | 0 | C | 22 | 0.0004 | rs1221010438 |
| 8 | c.239 + 150C>A | p.? | 4 | 2 | 0 | i1 | ND | C | This study | NA | |
| 9 | c.1012C>T | p.Gln338* | 4 | 2 | 0 | e6/J helix | ND | C | 23 | 0.0008 | rs1214983921 |
| 10 | c.206A>C | p.His69Pro | 4 | 2 | 0 | e1/A helix | ND | C | This study | NA | |
| 11 | c.563_566insTCCA | p.Gln189Hisfs*71 | 4 | 2 | 0 | e3/E helix | ND | C | 24 | NA | |
| 12 | c.595 + 12G>A | p.? | 3 | 1 | 1 | i3 | ND | C | 25 | 0.0088 | |
| 13 | c.1337G>A | p.Gly446Asp | 2 | 1 | 0 | e8/Cys pocket | ND | C | This study | NA | |
| 14 | c.1449_1451delGGT | p.Met483Ile | 2 | 1 | 0 | e9/C-term | ND | C | 26 | NA | |
| 15 | c.1398 + 2T>C | p.? | 2 | 1 | 0 | i8 | ND | C | 27 | 0.0008 | rs577022490 |
| 16 | c.593_595delAAG | p.Glu198del | 2 | 1 | 0 | e3/E helix | ND | C | This study | NA | |
| 17 | c.348G>C | p.Trp116Cys | 2 | 1 | 0 | e2/B-C loop | 3 | C | 18 | 0.0007 | rs772003869 |
| 18 | c.1201-9C>A | p.? | 2 | 1 | 0 | i7 | ND | C | 28 | NA | |
| 19 | c.1122-1G>C | p.? | 2 | 1 | 0 | i6 | ND | C | This study | NA | |
| 20 | c.1460T>G | p.Phe487Cys | 1 | 0 | 1 | e9/C-term | ND | C | This study | NA | |
| 21 | c.1398 + 5G>C | p.? | 1 | 0 | 1 | i8 | ND | C | 24 | NA | rs1563867837 |
| 22 | c.946G>A | p.Val316Met | 6 | 3 | 0 | e5/I helix | ND | NC | 13 | 0.0032 | rs375833424 |
| 23 | c.1466T>C | p.Leu489Ser | 4 | 2 | 0 | e9/C-term | ND | NC | 8 | 0.00039 | rs750428278 |
| 24 | c.427C>T | p.Arg143Trp | 4 | 2 | 0 | e3/C helix | 10 | NC | 10 | 0.016 | rs140336749 |
| 25 | c.281C>T | p.Pro94Leu | 2 | 1 | 0 | e2/B helix | 0-2 | NC | 7 | 0.0004 | rs104894070 |
| 26 | c.890C>T | p.Ala297Val | 1 | 0 | 1 | e5/H-I loop | 27 | NC | 11 | 0.0016 | rs375892072 |
| 27 | c.1066C>T ^a | p.Gln356* | 1 | 0 | 1 | e6/J-K loop | ND | NC ^a | 23 | 0.00005 | rs146124466 |
| 28 | c.955-3C>A ^a | p.? | 1 | 0 | 1 | i5 | ND | NC ^a | This study | NA | |
| 29 | c.1235G>T ^b | p.Arg412Leu | 1 | 0 | 1 | e8/K-L loop | ND | NC ^b | This study | NA | |
| 30 | c.799 + 2T>C ^b | p.? | 1 | 0 | 1 | i4 | ND | NC ^b | NA | NA | rs193922541 |

Novel mutations are in bold.

Abbreviations: MAF in GnomAD, minor allele frequency in Genome Aggregation Database; NA, variant does not have a gnomAD genomes entry; ND, Not done.

^aIn patient #76 (16), clinical and biochemical phenotype was NC-11 β OHD with compound heterozygosity for [c.1066C>T][c.955-3C>A] mutations in CYP11B1. c.1066C>T was reported to cause a truncated, nonfunctional protein. The enzymatic activity of c.955-3C>A variant has not been tested in vitro. Nonetheless, this variant was predicted to cause a milder gene defect and NC-11 β OHD phenotype was presumably related to the residual activity of this milder allele.

^bIn patient #72 (16), clinical and biochemical phenotype was NC-11 β OHD with compound heterozygosity for [c.1235G>T][c.799 + 2T] mutations in CYP11B1. The enzymatic activities of c.1235G>T and c.799 + 2T>C variants have not been tested in vitro. Nonetheless, at least 1 of these variants was predicted to cause a milder gene defect and NC-11 β OHD phenotype was presumably related to the residual activity of this milder allele.

were diagnosed earlier than NC-11 β OHD patients (median, 95%CI, and range: 1.33, 1.19 to 2.19, and 0.01 to 13 years *vs* 6.90, 5.3 to 11.2, 4.0 to 15.8 years, respectively; $P < 0.0001$). The bone age was more advanced in C-11 β OHD patients (bone age/chronological age ratio median, 95%CI, and range: 2.33, 2.40 to 3.74, and 0.80 to 15.00 for C-11 β OHD and 1.50, 1.27 to 1.78, and 1.01 to 2.18 years, respectively, for NC-11 β OHD; $P = 0.04$) (Fig. 1B). All of the C-11 β OHD and NC-11 β OHD patients in our study were on hydrocortisone replacement therapy. Hydrocortisone was initiated in NC-11 β OHD patients to control androgen excess. The median age of starting hydrocortisone treatment was 1.3 (range: 0.01 to 13) and 6.9 (range: 4.0 to 15.8) years in C- and NC-11 β OHD patients, respectively ($P < 0.0001$). Hydrocortisone treatment doses were not statistically different in C- and NC-11 β OHD patients (16.8 ± 5 and 14.4 ± 2.8 mg/m²/d in C- and NC-11 β OHD patients, respectively ($P = 0.14$)). Adult height SDS and corrected height SDS values were lower in C-11 β OHD ($n = 37$) than in NC-11 β OHD ($n = 5$) patients [-2.46 (range: -6.81 to -0.1) *vs* -1.32 (range -1.55 to -0.19), $P = 0.05$ and -1.5 (range: -4.87 to 1.64) *vs* -1.14 (range: -1.40 to -0.57), $P = 0.39$; respectively].

Plasma ACTH concentrations were higher in C-11 β OHD than in NC-11 β OHD patients [median (IQR): 142 (47 to 213) *vs* 10 (6 to 22) pmol/L, $P = 0.02$; normal range: 2.2 to 13.2 pmol/L]. Cortisol response to standard intravenous cosyntropin test (250 μ g) was 533 nmol/L (range: 450 to 687) in NC-11 β OHD patients ($n = 4$). Table 2 shows the comparison of adrenocortical steroid hormone measurements by LC-MS/MS between C- and NC-11 β OHD patients compared to the control group. The most significant elevations were in 11-deoxycortisol, DOC, 17OH-pregnenolone, and the adrenal androgen precursor (androstenedione) in decreasing order in 11 β OHD patients compared to controls. The most notable decrease was in cortisol, cortisone, and corticosterone concentrations in patients compared to controls. The 11-oxygenated androgens were also significantly lower in patients compared to controls. Cortisol, cortisone, 11-deoxycortisol, DOC, corticosterone, and DHEA concentrations were the main adrenal steroids, which differed between C- and NC-11 β OHD patients (Table 2). Among the steroid concentrations adjusted for cortisol, 11-deoxycortisol/cortisol, DOC/cortisol, and androstenedione/cortisol ratios were specific to discriminate C- and NC-11 β OHD patients, in decreasing order (Table 2, Fig. 2A, 2B, and 2F). The 17OH-pregnenolone/cortisol, 17OH-progesterone/cortisol, and DHEA/cortisol ratios were similar between C- and NC-11 β OHD patients (Table 2, Fig. 2C, 2D, and 2E). The ratio of 11-deoxycortisol/cortisol was significantly

different between C-11 β OHD patients with or without hypertension and NC-11 β OHD patients; [median (IQR): 42.3 (22.7-69.3) for C-11 β OHD with hypertension, 18.4 (8.15-37.7) for C-11 β OHD without hypertension and 0.35 (0.24-0.91) for NC-11 β OHD patients; $P < 0.0001$].

Discussion

This study describes the largest pediatric group, as well as the largest national cohort, of patients with C- and NC-11 β OHD and outlines the spectrum of this rare disease. Our findings demonstrate a concordance of clinical and metabolic profile of 11 β OHD patients with their molecular characteristics. Our study shows, in particular, the utility of the multiteroid profiling data that may predict the 11 β OHD phenotype (classic *vs* nonclassic).

Ambiguous genitalia was the leading sign of C-11 β OHD girls in our study, which is similar to 21OHD (34). On the other hand, we noticed that lack of salt-wasting delayed the time of diagnosis in C-11 β OHD boys further than boys with classic 21OHD (34) unless severe hypertension comes to clinical attention. This also applied to 10% of the 46,XX patients in our cohort with severe genital virilization (Prader 5) who had delayed diagnosis of C-11 β OHD and raised as males. Nevertheless, we found that C-11 β OHD patients were diagnosed before an age of 2.5 years, while the clinical presentation of a NC-11 β OHD before 5 years of age is unlikely. The absence of ambiguous genitalia in girls and the normal blood pressure were the main clinical differences between C- and NC-11 β OHD, explaining the delayed diagnosis in those with NC-11 β OHD. Instead, those patients were initially followed for other diagnoses such as premature pubarche, hirsutism, polycystic ovary syndrome, or NC-21OHD, similar to previous reports (6-11). Indeed, NC-11 β OHD should be an important diagnosis of exclusion in the clinical and biochemical work-up of disorders of androgen excess (3).

Short adult height is a feature of 11 β OHD patients (35). Our results showed that the adult height was approximately 16 cm (2.7 SD) shorter in C-11 β OHD and 6 cm (1.0 SD) shorter in NC-11 β OHD patients compared to healthy Turkish population (36,37). Impaired adult height maybe attributed to a delay in diagnosis and possible overreplacement with glucocorticoids. However, our patients with NC-11 β OHD were diagnosed significantly later than the ones with C-11 β OHD, and both groups were treated with the similar doses of hydrocortisone. We have also observed that the adult height is more compromised in 11 β OHD patients compared to 21OHD. The adult height of our patients with 11 β OHD were shorter than Turkish classic and NC-21OHD patients by

approximately 1.1 and 1.5 SD (6.6–9.0 cm), respectively (36,38,39). In line with this, bone age to chronological age ratio was >2 in almost all C-11 β OHD patients, and it was below 1.8 in almost all NC-11 β OHD patients. This may suggest that not only delayed diagnosis, but severity of the disease may contribute to impaired adult height in 11 β OHD. There were significantly increased 17OH-pregnenolone and androstenedione concentrations in both C- and NC-11 β OHD patients. The extent of 17OH-pregnenolone elevation was even more prominent than that of 17OH-progesterone. As 17OH-pregnenolone is the preferred substrate for the 17,20-lyase activity of CYP17A1, this allows for an increase in the biosynthesis of adrenal androgen precursors via the delta 5 pathway. Furthermore, the loss of CYP11B1 activity prevents the 11 β -hydroxylation of androstenedione thereby reducing the production of 11-oxygenated androgens and potentially leading to more androstenedione being available for the peripheral aromatization to estrogens. This may contribute to accelerated bone age, premature epiphyseal closure, and short adult height in these patients. This is the first study to confirm low 11-oxygenated androgens in this disorder due to loss of CYP11B1 activity and therefore the inability to convert A4 into 11OHA4.

Similar to some previous reports, one fifth of our patients were initially misdiagnosed and treated as 21OHD, the main differential diagnosis of 11 β OHD due to its clinical similarities and high 17OH-progesterone (40). The 11 β -hydroxylated products of 17OH-progesterone (21-deoxycortisol), androstenedione (11 β -hydroxyandrostenedione), and testosterone (11 β -hydroxytestosterone) have recently been shown to be increased in 21OHD patients (41,42). In contrast, our findings clearly show that low 21-deoxycortisol, 11 β -hydroxyandrostenedione, and 11 β -hydroxytestosterone can be regarded as a signature steroid profile of 11 β OHD during the differential diagnosis of 21OHD. High concentrations of 11-deoxycortisol, DOC, androstenedione, and low concentrations of cortisol, cortisone, and corticosterone should also be considered as a typical steroid fingerprint in 11 β OHD. Furthermore, 11-deoxycortisol, DOC, 17OH-pregnenolone, and androstenedione were the most markedly raised hormones in 11 β OHD patients compared to controls by 300, 150, 90, and 15 times, respectively. These results altogether suggest that simultaneous measurement of these steroids by LC-MS/MS are the most convenient method for diagnosis of 11 β OHD providing high accuracy and reducing the need for ACTH stimulation test. Similarly, diagnostic ratios calculated using gas chromatography mass spectrometry, which measures quantitative urinary steroid hormone profiles can readily establish the diagnosis of 11 β OHD. Indeed,

a single spot urine measuring 100* tetrahydrodeoxycortisol/ (tetrahydrocortisol + tetrahydrocortisone + 5 α -tetrahydrocortisol) measured in a single spot urine establishes the diagnosis of 11 β OHD with a high confidence in any age (43).

Unlike C-11 β OHD, baseline ACTH and stimulated cortisol levels were normal or near-normal in our patients with NC-11 β OHD. Nevertheless, there was a significant difference between basal measurements of 11-deoxycortisol, cortisol, DOC, corticosterone, cortisone, and DHEA concentrations, in descending order of significance, between C- and NC-11 β OHD patients. This suggests that steroidogenesis is less severely impaired in the glucocorticoid and mineralocorticoid pathways than in the adrenal sex steroid biosynthesis in NC-11 β OHD compared to C-11 β OHD. However, the high DOC, and adrenal androgens in NC-11 β OHD compared to controls indicate that these patients should have appropriate treatment and require surveillance to avoid long-term consequences of hyperandrogenism and potential hypertension.

Diagnostic specificity for the subgroups of 11 β OHD was increased using steroid concentrations in proportion to cortisol. Especially 11-deoxycortisol/cortisol ratio emerged as the best biochemical marker to differentiate C- and NC-11 β OHD patients. This ratio was around 75 times higher in C-11 β OHD than in NC-11 β OHD, compared to 5- to 6-fold difference in 11-deoxycortisol concentrations alone. No overlap was found in the 11-deoxycortisol/cortisol ratio, and the lower and upper 95% CIs were 27 to 40, 0.18 to 1.00, and 0.002 to 0.003 in C-11 β OHD and NC-11 β OHD patients and control groups, respectively. Furthermore, 11-deoxycortisol/cortisol ratio was significantly higher in the subset of C-11 β OHD patients with hypertension compared to those without. Our results support the concordance between genotype, clinical phenotype, and steroid metabolism in 11 β OHD patients.

We identified 9 novel and 21 previously reported mutations in our 102 children with 11 β OHD. There was an enrichment of four rare pathogenic alleles in CYP11B1 (p.Asn394Argfs*37, p.Leu299Pro, p.Thr318 = and p.Arg141*) in 58% of 11 β OHD patients, which represents a significant mutational load in the Turkish population. These 4 variants were the most common mutations previously described in Turkish patients with 11 β OHD (23,25). This may allow a more focused clinical genetic screening program for 11 β OHD to be established in Turkey.

There is no distinct demarcation between C- and NC-11 β OHD, and this is often a spectrum due to relative loss of CYP11B1 function as in the case of 21OHD CAH. The categorization of C- and NC-11 β OHD should be based on clinical, biochemical, and molecular grounds. There are less than 20 mutations associated

Table 2. Comparison of plasma adrenocortical steroid hormone measurements by LC-MS/MS between patients with classic and nonclassic 11 β -hydroxylase deficiency compared to the controls

| Pathway/steroid hormone (nmol/L) ^a | Classic 11 β OH Δ (n = 78), median (IQR) | Nonclassic 11 β OH Δ (n = 10), median (IQR) | Control (n = 210), median (IQR) | P-value | | Fold change (X) (pt/control) |
|---|---|--|---------------------------------|---------|---------------|------------------------------|
| | | | | C vs NC | Pt vs control | |
| Minerolocorticoid | | | | | | |
| Pregnenolone | 2.54 (0.31-6.82) | 4.99 (0.50-13.3) | 0.15 (0.03-0.50) | <0.0001 | <0.0001 | 5.26 |
| Progesterone | 1.71 (0.76-3.56) | 0.41 (0.25-1.33) | 0.19 (0.09-0.35) | 0.75 | 0.00 006 | 3.42 |
| 11-deoxycorticosterone | 25.7 (7.93-42.0) | 5.87 (1.27-7.27) | 0.12 (0.06-0.18) | <0.0001 | <0.0001 | 149.28 |
| Corticosterone | 0.66 (0.26-1.87) | 3.15 (0.98-14.7) | 14.5 (9.30-21.2) | 0.03 | <0.0001 | 0.15 |
| Aldosterone | 0.22 (0.08-0.36) | 0.13 (0.02-0.27) | 0.05 (0.02-0.11) | 0.48 | 0.0001 | 2.83 |
| Glucocorticoid | | | | | | |
| 17OH-pregnenolone | 9.93 (1.41-124.3) | 231 (31.9-448) | 0.66 (0.18-2.55) | <0.0001 | <0.0001 | 94.16 |
| 17OH-progesterone | 6.43 (2.83-12.4) | 2.38 (2.11-3.74) | 0.90 (0.42-1.41) | 0.0009 | <0.0001 | 10.07 |
| 11-deoxycortisol | 348 (138.6-500) | 68.2 (47.2-100) | 0.72 (0.37-1.26) | <0.0001 | <0.0001 | 314.44 |
| Cortisol | 8.91 (5.83-14.9) | 190 (158-254) | 285 (199-427) | 0.06 | <0.0001 | 0.12 |
| Cortisone | 0.60 (0.25-1.77) | 61.7 (46.7-140.9) | 67.1 (49.8-81.9) | 0.003 | <0.0001 | 0.36 |
| 21-Deoxycortisol | 0.05 (0.02-0.20) | 0.08 (0.02-0.46) | 0.11 (0.05-0.28) | 0.71 | 0.15 | 1.03 |
| Androgen | | | | | | |
| DHEA | 4.61 (0.97-10.8) | 37.1 (9.82-52.0) | 3.64 (0.79-8.50) | <0.0001 | 0.0001 | 2.11 |
| Androstenedione | 19.8 (4.36-32.1) | 8.96 (7.01-24.3) | 0.87 (0.38-2.51) | <0.0001 | <0.0001 | 13.24 |
| Testosterone | 2.35 (0.79-7.35) | 1.38 (0.76-4.30) | 0.31 (0.13-1.24) | 0.94 | 0.002 | 1.98 |
| Androsterone | 4.46 (1.25-10.7) | 5.61 (2.54-6.70) | 0.41 (0.10-1.64) | 0.02 | <0.0001 | 5.45 |
| 11 β -hydroxyandrostenedione | 0.76 (0.39-1.85) | 0.59 (0.39-1.25) | 27.8 (17.0-42.3) | 0.007 | <0.0001 | 0.04 |
| 11 β -hydroxytestosterone | 0.06 (0.03-0.19) | 0.06 (0.03-0.16) | 0.16 (0.06-0.39) | 0.31 | 0.0001 | 0.51 |
| Steroid hormone ratios | | | | | | |
| 11-deoxycortisol/cortisol | 27.75 (12.22-47.92) | 0.35 (0.24-0.91) | 0.002 (0.001-0.003) | <0.0001 | <0.0001 | 10 849 |
| 11-deoxycorticosterone/cortisol | 2.43 (0.89-3.60) | 0.02 (0.01-0.03) | 0.0003 (0.0001-0.0005) | <0.0001 | <0.0001 | 5561 |
| 17OH-pregnenolone/cortisol | 1.38 (0.18-18.18) | 1.05 (0.29-2.66) | 0.002 (0.0005-0.008) | <0.0001 | <0.0001 | 3228 |
| 17OH-progesterone/cortisol | 0.68 (0.26-1.35) | 0.01 (0.009-0.04) | 0.002 (0.001-0.004) | <0.0001 | <0.0001 | 343 |
| DHEA/cortisol | 0.38 (0.11-1.18) | 0.14 (0.08-0.16) | 0.01 (0.002-0.020) | <0.0001 | <0.0001 | 55 |
| Androstenedione/cortisol | 1.20 (0.41-2.97) | 0.03 (0.02-0.11) | 0.002 (0.001-0.006) | <0.0001 | <0.0001 | 399 |

Abbreviations: C, classic; DHEA, dehydroepiandrosterone; IQR, interquartile range; NC, nonclassic; Pt, patient.

^aTo convert nmol/L to ng/mL, divide by 2.77 for aldosterone, 2.89 for corticosterone, 3.03 for 11-deoxycorticosterone, 3.10 for pregnenolone, 3.02 for 17 α -hydroxyprogesterone, 3.18 for progesterone, 3.01 for 17 α -hydroxypregnenolone, 2.88 for 21-deoxycortisol and 11-deoxycortisol, 2.75 for cortisol, 2.77 for cortisone, 3.47 for DHEA, 2.71 for DHEAS, 3.49 for androstenedione and androsterone, 3.47 for testosterone, 3.31 for 11 β -hydroxyandrostenedione, and 3.28 for 11 β -hydroxytestosterone.

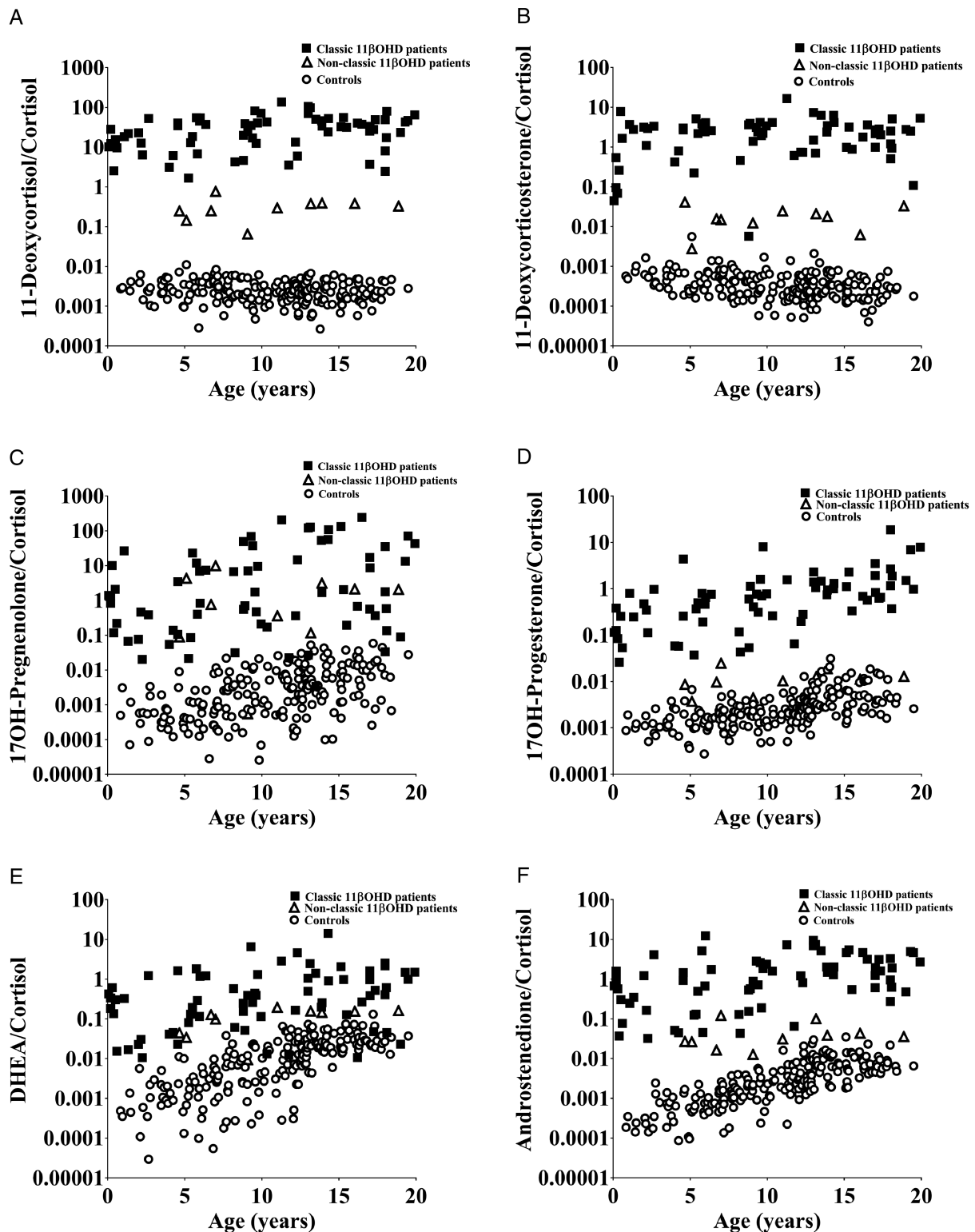


Figure 2. Steroid ratios across ages in patients with classic and nonclassical 11 β -hydroxylase deficiency. Data are shown as linear regressions of steroids normalized to cortisol concentrations as a function of age (in years). All ratios showed a significant difference between patients and controls. 11-deoxycortisol/cortisol (A), 11-deoxycorticosterone/cortisol (B) and androstenedione/cortisol (F) ratios, but not 17OH-pregnenolone/cortisol (C), 17OH-progesterone/cortisol (D), and DHEA/cortisol (E) ratios could discriminate classic and nonclassical 11 β -hydroxylase deficiency.

with NC-11 β OHD in the literature (3,11). A residual CYP11B1 activity of above 10% to 25% is associated with a clinical phenotype of NC-11OHD similar to

NC-21OHD or mild simple virilizing 21OHD (3,11). As in 21OHD patients (44), we observed in our patients with NC-11 β OHD that a compound heterozygosity for

1 mild and 1 severe mutation in *CYP11B1* had a less severe clinical presentation than those patients with 2 severe mutations (ie, the phenotype was determined by the least severe mutation).

The main limitation of our study was the retrospective nature of the study. As we collected the clinical and biochemical characteristics retrospectively from medical files, some data regarding initial presentation were incomplete. Another limitation was the relatively small subgroup size of NC-11 β OHD patients. Despite the large number of study individuals, comparisons of some characteristics between C- and NC-11 β OHD patients should be interpreted with caution.

In conclusion, 11 β OHD is an important diagnosis of exclusion in hypertension and androgen excess states in childhood. Nevertheless, it is not always straightforward to establish the diagnosis due to its broad clinical spectrum depending on the genotype. Especially NC-11 β OHD can escape from clinical attention due to relatively mild clinical presentations and near-normal first-line adrenal function tests. However, the retained enzyme activity of *CYP11B1* showed a good correlation with LC-MS/MS-based steroid profiles, which enable the diagnosis and subtyping of 11 β OHD.

Acknowledgments

We are deeply grateful to the patients and families without whom this study could not have been performed.

Financial Support: This work has been supported by the Medical Research Council of Marmara University (Project Grant SAG-A-120418-0152, TG).

Author contributions: TG, MY, and AB designed the study. TG, MY, EI, ZYA, MK, MNO, FB, AU, MB, CK, HO, FD, AC, EU, AA, HD, TC, FD, GC, ST, and AB recruited and clinically characterized the patients. TG, KHS, AY, and GH conducted and analyzed biochemical and LC-MS/MS measurements. TG, ZYA, and TB performed and analyzed the Sanger sequencing of *CYP11B1*. TG, MY, ZYA, HF, and AB prepared the draft manuscript. All authors contributed to the discussion of results and edited and approved the final manuscript.

Additional Information

Correspondence: Tulay Guran, Marmara University School of Medicine, Department of Pediatric Endocrinology and Diabetes Fevzi Cakmak Mh. Mimar Sinan Cd. No 41. 34899 Ustkaýnarca/Pendik Istanbul, Turkey. Email: tulay.guran@marmara.edu.tr

Disclosures: The authors have nothing to disclose.

Data Availability: Some data sets analyzed during the current study are included in the data repositories listed in references.

References

1. El-Maouche D, Arlt W, Merke DP. Congenital adrenal hyperplasia. *Lancet*. 2017;390(10108):2194-2210.
2. Khattab A, Haider S, Kumar A, et al. Clinical, genetic, and structural basis of congenital adrenal hyperplasia due to 11 β -hydroxylase deficiency. *Proc Natl Acad Sci U S A*. 2017;114(10):E1933-E1940.
3. Bulsari K, Falhammar H. Clinical perspectives in congenital adrenal hyperplasia due to 11 β -hydroxylase deficiency. *Endocrine*. 2017;55(1):19-36.
4. White PC, Curnow KM, Pascoe L. Disorders of steroid 11 beta-hydroxylase isozymes. *Endocr Rev*. 1994;15(4):421-438.
5. Gran T, Tezel B, akır M, et al. Neonatal screening for congenital adrenal hyperplasia in Turkey: outcomes of extended pilot study in 241 083 infants. *J Clin Res Pediatr Endocrinol*. 2020;12(3):287-294.
6. Joehrer K, Geley S, Strasser-Wozak EM, et al. *CYP11B1* mutations causing non-classic adrenal hyperplasia due to 11 beta-hydroxylase deficiency. *Hum Mol Genet*. 1997;6(11):1829-1834.
7. Krone N, Grischuk Y, Mller M, et al. Analyzing the functional and structural consequences of two point mutations (P94L and A368D) in the *CYP11B1* gene causing congenital adrenal hyperplasia resulting from 11-hydroxylase deficiency. *J Clin Endocrinol Metab*. 2006;91(7):2682-2688.
8. Peters CJ, Nugent T, Perry LA, et al. Cosegregation of a novel homozygous *CYP11B1* mutation with the phenotype of non-classical congenital adrenal hyperplasia in a consanguineous family. *Horm Res*. 2007;67(4):189-193.
9. Parajes S, Loidi L, Reisch N, et al. Functional consequences of seven novel mutations in the *CYP11B1* gene: four mutations associated with nonclassic and three mutations causing classic 11{beta}-hydroxylase deficiency. *J Clin Endocrinol Metab*. 2010;95(2):779-788.
10. Reisch N, Hgler W, Parajes S, et al. A diagnosis not to be missed: nonclassic steroid 11 β -hydroxylase deficiency presenting with premature adrenarche and hirsutism. *J Clin Endocrinol Metab*. 2013;98(10):E1620-E1625.
11. Mooij CF, Parajes S, Rose IT, et al. Characterization of the molecular genetic pathology in patients with 11 β -hydroxylase deficiency. *Clin Endocrinol (Oxf)*. 2015;83(5):629-635.
12. Wang D, Wang J, Tong T, Yang Q. Non-classical 11 β -hydroxylase deficiency caused by compound heterozygous mutations: a case study and literature review. *J Ovarian Res*. 2018;11(1):82.
13. Zacharieva S, Robeva R, Andonova S, et al. Long-term follow-up of a female patient with non-classical 11 β -hydroxylase deficiency and two novel mutations in *CYP11B1*. *Gynecol Endocrinol*. 2019;35(1):23-27.
14. Guran T, Kara C, Yildiz M, et al. Revisiting classical 3 β -hydroxysteroid dehydrogenase 2 deficiency: lessons from 31 pediatric cases. *J Clin Endocrinol Metab*. 2020;105(3):dgaa022.
15. Nguyen HH, Eiden-Plach A, Hannemann F, et al. Phenotypic, metabolic, and molecular genetic characterization of six patients with congenital adrenal hyperplasia caused by novel mutations in the *CYP11B1* gene. *J Steroid Biochem Mol Biol*. 2016;155(Pt A):126-134.
16. Yildiz M, Isik E, Abali ZY, et al. Data from: Clinical and hormonal profile correlates with molecular characteristics in patients with 11 β -hydroxylase deficiency—supplemental table S1. Figshare. Deposited March 3, 2021. doi:10.6084/m9.figshare.14153372
17. Helmborg A, Ausserer B, Kofler R. Frame shift by insertion of 2 basepairs in codon 394 of *CYP11B1* causes congenital adrenal hyperplasia due to steroid 11 beta-hydroxylase deficiency. *J Clin Endocrinol Metab*. 1992;75(5):1278-1281.

18. Krone N, Riepe FG, Götze D, et al. Congenital adrenal hyperplasia due to 11-hydroxylase deficiency: functional characterization of two novel point mutations and a three-base pair deletion in the CYP11B1 gene. *J Clin Endocrinol Metab.* 2005;**90**(6):3724-3730.
19. Skinner CA, Rumsby G, Honour JW. Single strand conformation polymorphism (SSCP) analysis for the detection of mutations in the CYP11B1 gene. *J Clin Endocrinol Metab.* 1996;**81**(6):2389-2393.
20. Sólyom J, Cholnoky P, Helmuth D, et al. A steroid-11-beta-hydroxylase defektus diagnosztikája [Diagnosis of steroid-11-beta hydroxylase deficiency]. *Orv Hetil.* 1986;**127**(36):2171-2175.
21. Polat S, Kulle A, Karaca Z, et al. Characterisation of three novel CYP11B1 mutations in classic and non-classic 11 β -hydroxylase deficiency. *Eur J Endocrinol.* 2014;**170**(5):697-706.
22. Geley S, Kapelari K, Jöhrer K, et al. CYP11B1 mutations causing congenital adrenal hyperplasia due to 11 beta-hydroxylase deficiency. *J Clin Endocrinol Metab.* 1996;**81**(8):2896-2901.
23. Curnow KM, Slutsker L, Vitek J, et al. Mutations in the CYP11B1 gene causing congenital adrenal hyperplasia and hypertension cluster in exons 6, 7, and 8. *Proc Natl Acad Sci U S A.* 1993;**90**(10):4552-4556.
24. Kandemir N, Yilmaz DY, Gonc EN, et al. Novel and prevalent CYP11B1 gene mutations in Turkish patients with 11- β hydroxylase deficiency. *J Steroid Biochem Mol Biol.* 2017;**165**(Pt A):57-63.
25. Kim JH, Park G, Kim SY, Bae HY. Thyrotoxic periodic paralysis with Graves' disease leading to the discovery of a hidden nonclassical 11 β hydroxylase deficiency. *Intern Med.* 2013;**52**(1):85-88.
26. Baş F, Toksoy G, Ergun-Longmire B, et al. Prevalence, clinical characteristics and long-term outcomes of classical 11 β -hydroxylase deficiency (11BOHD) in Turkish population and novel mutations in CYP11B1 gene. *J Steroid Biochem Mol Biol.* 2018;**181**:88-97.
27. Breil T, Yakovenko V, Inta I, et al. Typical characteristics of children with congenital adrenal hyperplasia due to 11 β -hydroxylase deficiency: a single-centre experience and review of the literature. *J Pediatr Endocrinol Metab.* 2019;**32**(3):259-267.
28. Zhang M, Liu Y, Sun S, et al. A prevalent and three novel mutations in CYP11B1 gene identified in Chinese patients with 11-beta hydroxylase deficiency. *J Steroid Biochem Mol Biol.* 2013;**133**:25-29.
29. Adzhubei I, Jordan DM, Sunyaev SR. Predicting functional effect of human missense mutations using PolyPhen-2. *Curr Protoc Hum Genet.* 2013;Chapter 7:Unit7.20. doi:10.1002/0471142905.hg0720s76
30. Sim NL, Kumar P, Hu J, Henikoff S, Schneider G, Ng PC. SIFT web server: predicting effects of amino acid substitutions on proteins. *Nucleic Acids Res.* 2012;**40**(Web Server issue):W452-W457.
31. Schwarz JM, Cooper DN, Schuelke M, Seelow D. MutationTaster2: mutation prediction for the deep-sequencing age. *Nat Methods.* 2014;**11**(4):361-362.
32. Choi Y, Chan AP. PROVEAN web server: a tool to predict the functional effect of amino acid substitutions and indels. *Bioinformatics.* 2015;**31**(16):2745-2747.
33. Abecasis GR, Auton A, Brooks LD, et al; 1000 Genomes Project Consortium. An integrated map of genetic variation from 1092 human genomes. *Nature.* 2012;**491**(7422):56-65.
34. Speiser PW, Auchus RJ, Merke DP, Miller WL, White PC. Congenital adrenal hyperplasia due to steroid 21-hydroxylase deficiency: an endocrine society clinical practice guideline. *J Clin Endocrinol Metab.* 2019;**104**(6):1928.
35. Hochberg Z, Schechter J, Benderly A, Leiberman E, Rosler A. Growth and pubertal development in patients with congenital adrenal hyperplasia due to 11-beta-hydroxylase deficiency. *Am J Dis Child.* 1985;**139**(8):771-776.
36. Günöz H, Bundak R, Furman A, et al. Z-score reference values for height in Turkish children aged 6 to 18 years. *J Clin Res Pediatr Endocrinol.* 2014;**6**(1):28-33.
37. Neyzi O, Bundak R, Gökçay G, et al. Reference values for weight, height, head circumference, and body mass index in Turkish children. *J Clin Res Pediatr Endocrinol.* 2015;**7**(4):280-293.
38. Savaş-Erdeve Ş, Çetinkaya S, Abalı ZY, et al. Clinical, biochemical and genetic features with nonclassical 21-hydroxylase deficiency and final height. *J Pediatr Endocrinol Metab.* 2017;**30**(7):759-766.
39. Aycan Z, Akbuğa S, Cetinkaya E, et al. Final height of patients with classical congenital adrenal hyperplasia. *Turk J Pediatr.* 2009;**51**(6):539-544.
40. Tonetto-Fernandes V, Lemos-Marini SH, Kuperman H, Ribeiro-Neto LM, Verreschi IT, Kater CE. Serum 21-deoxycortisol, 17-hydroxyprogesterone, and 11-deoxycortisol in classic congenital adrenal hyperplasia: clinical and hormonal correlations and identification of patients with 11beta-hydroxylase deficiency among a large group with alleged 21-hydroxylase deficiency. *J Clin Endocrinol Metab.* 2006;**91**(6):2179-2184.
41. Turcu AF, Nanba AT, Chomic R, et al. Adrenal-derived 11-oxygenated 19-carbon steroids are the dominant androgens in classic 21-hydroxylase deficiency. *Eur J Endocrinol.* 2016;**174**(5):601-609.
42. Miller WL. Congenital adrenal hyperplasia: time to replace 17OHP with 21-deoxycortisol. *Horm Res Paediatr.* 2019;**91**(6):416-420.
43. Krone N, Hughes BA, Lavery GG, Stewart PM, Arlt W, Shackleton CH. Gas chromatography/mass spectrometry (GC/MS) remains a pre-eminent discovery tool in clinical steroid investigations even in the era of fast liquid chromatography tandem mass spectrometry (LC/MS/MS). *J Steroid Biochem Mol Biol.* 2010;**121**(3-5):496-504.
44. Weintrob N, Brautbar C, Pertzlan A, et al. Genotype-phenotype associations in non-classical steroid 21-hydroxylase deficiency. *Eur J Endocrinol.* 2000;**143**(3):397-403.